

## Research



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## Animal behaviour

## Microplastic leachates impair behavioural vigilance and predator avoidance in a temperate intertidal gastropod

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Microplastics are a ubiquitous source of contaminations in marine ecosystems, and have major implications for marine life. Much effort has been devoted to assessing the various effects of microplastics on marine life. No evidence exists, however, on the effects of microplastic leachates on chemically mediated predator–prey interactions and the ability of prey to detect and avoid its predator. This study shows that microplastic leachates have direct biological effects by disturbing the behavioural response of the intertidal gastropod *Littorina littorea* to the presence of *Carcinus maenas* chemical cues, hence increasing their vulnerability to predation. Leachates from virgin and beached pellets respectively impaired and inhibited *L. littorea* vigilance and anti-predator behaviours. These results suggest that the biological effects from microplastic leachates may have major implications for marine ecosystems on taxa that rely on chemosensory cues to escape predation.

## 1. Introduction

Plastics are a major source of global marine pollution [1]. They have conspicuous effects such as the deposition of beached debris [2] and the entanglement of marine fauna [3]. Microplastics (i.e. plastic particles less than 5 mm) are a more recent and pernicious source of pollution, persistent and nearly ubiquitous in marine systems [1]. One of the greatest concerns about microplastics in marine environments is their effect on marine organisms [3]. Microplastics are a vector of chemical pollutants adsorbed onto their surface. Their ingestion by organisms as small as zooplankton prompts the desorption of these chemicals which cause adverse effects [4]. They also accumulate into the tissues and organs, hence cascade through the food chain [5]. The effects of microplastics through the release in the environment of additives used in their manufacture [6] and the contaminants that adsorb and accumulate onto their surface [7] have been far less studied. They were essentially inferred through experiments assessing the toxicity of leachates from new plastic consumer products to aquatic invertebrates [8]. The potential effect of microplastic leachates is critical as most plastic contamination in the ocean is made of microplastics, which tend to accumulate more persistent pollutants than large debris [9].

A key component of microplastic pollution is raw resin pellets used in the manufacture of plastic products [10]. These pellets are found on beaches worldwide, and accumulate various types of persistent pollutants [11]. However, only a few studies have assessed the toxicity of leachate from plastic pellets, and found high toxicity to the embryonic development of marine invertebrates [12,13]. So far, no evidence exists on the potential effect of plastic pellet leachate on predator–prey interactions, in particular on the ability of prey to detect and avoid its predator. These species-specific behaviours are typically mediated by chemical cues [14], whose detection is compromised by chemical pollution [15]. This study assessed experimentally whether prey vigilance and

**Table 1.** Statistical assessment of the differences between experimental treatments for each behavioural variable.

behavioural variable	treatment	statistic	d.f.	<i>p</i>
righting time	overall (Kruskal – Wallis)	$H = 196.7$	5	<0.001
	SW versus SW <sub>CC</sub>	$U = 0$	2	<0.001
	SW versus SW <sub>VP</sub>	$U = 1088$	2	>0.05
	SW versus SW <sub>BP</sub>	$U = 830.5$	2	>0.05
	SW versus SW <sub>CC-VP</sub>	$U = 126.5$	2	<0.001
	SW versus SW <sub>CC-BP</sub>	$U = 966.5$	2	>0.05
	SW <sub>CC</sub> versus SW <sub>VP</sub>	$U = 0$	2	<0.001
	SW <sub>CC</sub> versus SW <sub>BP</sub>	$U = 0$	2	<0.001
	SW <sub>CC</sub> versus SW <sub>CC-VP</sub>	$U = 133.5$	2	<0.001
	SW <sub>CC</sub> versus SW <sub>CC-BP</sub>	$U = 0$	2	<0.001
	SW <sub>VP</sub> versus SW <sub>BP</sub>	$U = 1012$	2	>0.05
	SW <sub>VP</sub> versus SW <sub>CC-VP</sub>	$U = 139$	2	<0.001
	SW <sub>VP</sub> versus SW <sub>CC-BP</sub>	$U = 1191$	2	>0.05
	SW <sub>BP</sub> versus SW <sub>CC-VP</sub>	$U = 76$	2	<0.001
	SW <sub>BP</sub> versus SW <sub>CC-BP</sub>	$U = 1048$	2	>0.05
	SW <sub>CC-VP</sub> versus SW <sub>CC-BP</sub>	$U = 0$	2	<0.001
time to explore	overall (Kruskal – Wallis)	$H = 209.2$	5	<0.001
	SW versus SW <sub>CC</sub>	$U = 0$	2	<0.001
	SW versus SW <sub>VP</sub>	$U = 1073$	2	>0.05
	SW versus SW <sub>BP</sub>	$U = 1240$	2	>0.05
	SW versus SW <sub>CC-VP</sub>	$U = 10$	2	<0.001
	SW versus SW <sub>CC-BP</sub>	$U = 988.5$	2	>0.05
	SW <sub>CC</sub> versus SW <sub>VP</sub>	$U = 0$	2	<0.001
	SW <sub>CC</sub> versus SW <sub>BP</sub>	$U = 0$	2	<0.001
	SW <sub>CC</sub> versus SW <sub>CC-VP</sub>	$U = 7$	2	<0.001
	SW <sub>CC</sub> versus SW <sub>CC-BP</sub>	$U = 0$	2	<0.001
	SW <sub>VP</sub> versus SW <sub>BP</sub>	$U = 1043$	2	>0.05
	SW <sub>VP</sub> versus SW <sub>CC-VP</sub>	$U = 10$	2	<0.001
	SW <sub>VP</sub> versus SW <sub>CC-BP</sub>	$U = 1177$	2	>0.05
	SW <sub>BP</sub> versus SW <sub>CC-VP</sub>	$U = 0$	2	<0.001
	SW <sub>BP</sub> versus SW <sub>CC-BP</sub>	$U = 982$	2	>0.05
	SW <sub>CC-VP</sub> versus SW <sub>CC-BP</sub>	$U = 0$	2	<0.001
skioptic withdrawal	overall (Kruskal – Wallis)	$H = 203.3$	5	<0.0001
	SW versus SW <sub>CC</sub>	$U = 0$	2	<0.0001
	SW versus SW <sub>VP</sub>	$U = 1046$	2	>0.05
	SW versus SW <sub>BP</sub>	$U = 1208$	2	>0.05
	SW versus SW <sub>CC-VP</sub>	$U = 129$	2	<0.0001
	SW versus SW <sub>CC-BP</sub>	$U = 1196$	2	>0.05
	SW <sub>CC</sub> versus SW <sub>VP</sub>	$U = 0$	2	<0.0001
	SW <sub>CC</sub> versus SW <sub>BP</sub>	$U = 0$	2	<0.0001
	SW <sub>CC</sub> versus SW <sub>CC-VP</sub>	$U = 359.5$	2	<0.0001
	SW <sub>CC</sub> versus SW <sub>CC-BP</sub>	$U = 0$	2	<0.0001
	SW <sub>VP</sub> versus SW <sub>BP</sub>	$U = 1007$	2	<0.0001
	SW <sub>VP</sub> versus SW <sub>CC-VP</sub>	$U = 182.5$	2	<0.0001
	SW <sub>VP</sub> versus SW <sub>CC-BP</sub>	$U = 1124$	2	>0.05
	SW <sub>BP</sub> versus SW <sub>CC-VP</sub>	$U = 121$	2	<0.0001

(Continued.)

Table 1. (Continued.)

behavioural variable	treatment	statistic	d.f.	p
avoidance response	SW <sub>BP</sub> versus SW <sub>CC-BP</sub>	$U = 1160$	2	>0.05
	SW <sub>CC-VP</sub> versus SW <sub>CC-BP</sub>	$U = 171.5$	2	<0.0001
	overall (Kruskal–Wallis)	$H = 201.3$	5	<0.001
	SW versus SW <sub>CC</sub>	$U = 0$	2	<0.001
	SW versus SW <sub>VP</sub>	$U = 1127$	2	>0.05
	SW versus SW <sub>BP</sub>	$U = 747.5$	2	>0.05
	SW versus SW <sub>CC-VP</sub>	$U = 0$	2	<0.001
	SW versus SW <sub>CC-BP</sub>	$U = 1019$	2	>0.05
	SW <sub>CC</sub> versus SW <sub>VP</sub>	$U = 0$	2	<0.001
	SW <sub>CC</sub> versus SW <sub>BP</sub>	$U = 0$	2	<0.001
	SW <sub>CC</sub> versus SW <sub>CC-VP</sub>	$U = 64.5$	2	<0.001
	SW <sub>CC</sub> versus SW <sub>CC-BP</sub>	$U = 0$	2	<0.001
	SW <sub>VP</sub> versus SW <sub>BP</sub>	$U = 976$	2	>0.05
	SW <sub>VP</sub> versus SW <sub>CC-VP</sub>	$U = 4.5$	2	<0.001
	SW <sub>VP</sub> versus SW <sub>CC-BP</sub>	$U = 1170$	2	>0.05
	SW <sub>BP</sub> versus SW <sub>CC-VP</sub>	$U = 10$	2	<0.001
SW <sub>BP</sub> versus SW <sub>CC-BP</sub>	$U = 1146$	2	>0.05	
SW <sub>CC-VP</sub> versus SW <sub>CC-BP</sub>	$U = 9$	2	<0.001	

antipredator behaviour of the intertidal gastropod *Littorina littorea* is impaired by leachates from virgin and beached plastic pellets.

## 2. Material and methods

*Littorina littorea* collected in August 2016 from mid-intertidal rock-pools at the Pointe du Noirda (50°49'45.15 N, 1°35'19.366 E)—a reef typical of the French coast of the eastern English Channel—were exposed to a factorial treatment of normal versus microplastic leachates seawater and the presence of chemical cues from the predatory crab *Carcinus maenas*. Snails (mean shell length  $10.1 \pm 1.2$  mm) were acclimated without food for 24 h in the laboratory in 120 l tanks of running natural seawater at 18°C, and each individual was only used once.

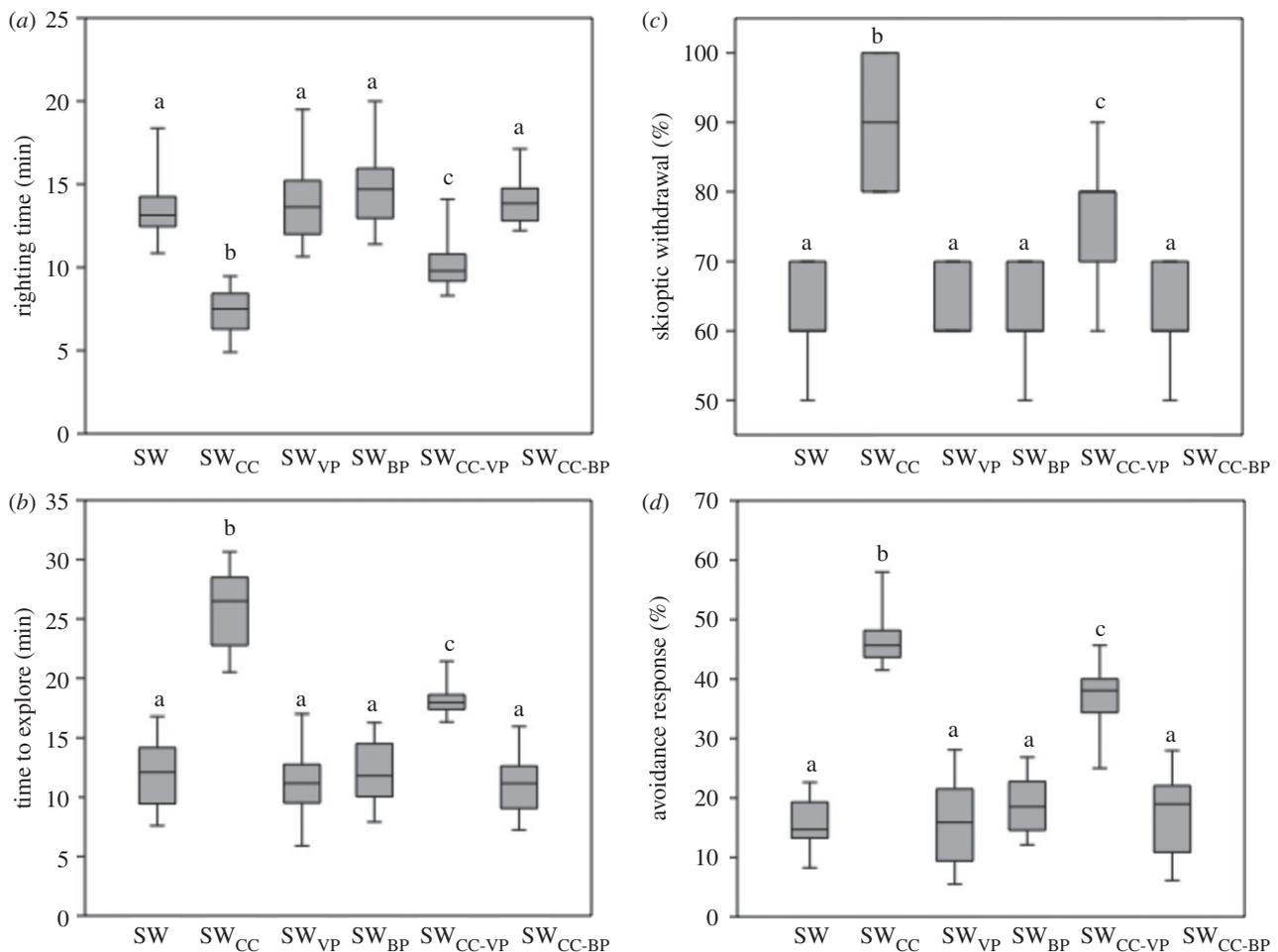
Microplastic leachate seawater was prepared from commercially available virgin polypropylene pellets (SW<sub>VP</sub>), and from beached pellets (SW<sub>BP</sub>) haphazardly collected from the high-tide mark sediment surface of beaches surrounding the reef. Pellets (diameter 3.3–4.7 mm) were mixed with seawater at a concentration of 20 ml of pellets per litre—a concentration one order of magnitude lower than those that impaired the embryonic development of mussels and sea urchins [12,13], which is not uncommon in the sampling area—and left for 24 h before the beginning of the behavioural assays. Crab cue seawater (SW<sub>CC</sub>) was prepared through the addition of an individual *C. maenas* (carapace width  $80.3 \pm 3.3$  mm) into 5 l of aerated seawater for a 48 h period prior to the trial. Crabs were previously fed *L. littorea* for a period of 10 days in the laboratory to stimulate predator-induced alarm responses [16]. Mixtures of SW<sub>CC</sub> and SW<sub>VP</sub> (SW<sub>CC-VP</sub>) and SW<sub>CC</sub> and SW<sub>BP</sub> (SW<sub>CC-BP</sub>) were prepared at a 1:1 ratio using crab cues and microplastic leachates that were twice as concentrated as in SW<sub>CC</sub> and SW<sub>VP</sub> to ensure they were present at the same concentrations than in other experiments.

Individual snails were placed in 500 ml borosilicate beakers filled with 300 ml of control and cue seawater for a 30 min

acclimation period. Three measures of prey vigilance and predator detection were investigated [17]. First, snails were dislodged using tissue forceps, placed onto their dorsal surface, and the righting time needed to regain contact with the substrate was taken in each treatment ( $N = 50$ ). Second, snails were dislodged, immediately placed into a beaker filled with the same water treatment, and the time to explore determined as the time taken by snails to re-emerge from their shell, extend their tentacles and began to actively crawl along the substrate ( $N = 50$ ). Finally, the skioptic (i.e. shadow-induced) withdrawal response was measured by passing a shadow over a snail actively crawling along the substrate. A 35 W halogen light was placed 50 cm above 500 ml borosilicate beakers filled with 300 ml of control and cue seawater. A 20 cm diameter black PVC piece was placed above experimental beakers for 5 s. Withdrawal was successful if the snail completely retracted into their shell within the 5 s interval. Each snail was assessed 10 times at a 1 min interval and withdrawal quantified as the percentage of individuals that successfully withdrew ( $N = 50$ ).

The percentage of time spent in a refuge or either above or at the water surface was used as a measure of avoidance [18]. These trials were performed in glass aquaria (15 × 15 cm, filled with 1100 ml of control and cue seawater) where PVC tubes (length 8 cm, diameter 5 cm) cut in half and glued down onto the centre of the aquarium created a refuge. Snails were placed next to the refuge, and left to acclimatize for 30 min. Their position was subsequently noted every 5 s for an hour after the addition of 100 ml of cue or non-cue water.

Individual snails were randomly exposed to control seawater (SW) and cue seawater (SW<sub>VP</sub>, SW<sub>BP</sub>, SW<sub>CC</sub>, SW<sub>CC-VP</sub> and SW<sub>CC-BP</sub>) to avoid the effect of trial order on responses. Behavioural traits were also investigated randomly. As circatidal rhythms could be a confounding factor, all experiments were conducted within a 2-h window before and after high-tide. As none of the four behavioural measures conformed to the normality assumption (Kolmogorov–Smirnov test,  $p < 0.05$ ), multiple comparisons between treatments were conducted using the Kruskal–Wallis



**Figure 1.** Behavioural responses of *Littorina littorea* to control seawater (SW), seawater conditioned with *Carcinus maenas* cues (SW<sub>CC</sub>), leachates from virgin (SW<sub>VP</sub>) and beached (SW<sub>BP</sub>) polypropylene pellets, and a 1:1 mixture of SW<sub>CC</sub> and SW<sub>VP</sub> (SW<sub>CC-VP</sub>) and SW<sub>CC</sub> and SW<sub>BP</sub> (SW<sub>CC-BP</sub>). The letters 'a', 'b' and 'c' indicate significantly distinct groups of measurements. The box represents the 25–75 quartiles, the median is shown with a horizontal line inside the box and the vertical lines indicate the minimal and maximal values.

test, followed by a Mann–Whitney *U*-test to check for significant pairwise differences.

### 3. Results and discussion

*Carcinus maenas* cue had a significant effect (table 1) on vigilance and antipredator responses of *L. littorea*. Snails exposed to crab cue (SW<sub>CC</sub>) significantly decreased their righting time compared with snails in control seawater (SW, figure 1a). Similarly, snails exposed to SW<sub>CC</sub> took significantly longer to emerge from their shells than following SW exposure (figure 1b) and elicited a full body withdrawal more often when presented a passing shadow that did snails in SW (figure 1c). *Littorina littorea* exhibited antipredator behaviour significantly more frequently in SW<sub>CC</sub> than in SW (figure 1d). These data demonstrate that *L. littorea* significantly increased their vigilance and antipredator responses following exposure to SW<sub>CC</sub> [19,20].

Leachates from both virgin (SW<sub>VP</sub>) and beached pellets (SW<sub>BP</sub>) did not elicit any significant change in snail behaviour in SW (figure 1). In turn, snails exposed to SW<sub>CC-VP</sub> significantly decreased vigilance and antipredator responses compared to those exposed to SW, SW<sub>VP</sub> and SW<sub>BP</sub> (table 1). These responses were, however, less pronounced than following an exposure to crab cue (table 1). Finally, *L. littorea* did not

significantly modify their behavioural responses when exposed to SW<sub>CC-BP</sub> compared with SW, SW<sub>VP</sub> and SW<sub>BP</sub> (table 1, figure 1). These results suggest that leachates from virgin and beached microplastic pellets did not affect *L. littorea* neuromuscular performance, while they respectively impaired and inhibited their ability to respond to predator cues through a decrease in their chemosensory abilities.

The alteration observed in *L. littorea* responses likely results from different contaminants and concentrations in the leachates from virgin and beached pellets. Virgin pellets essentially contain plastic additives (e.g. ultraviolet stabilizers and antimicrobials) that are adsorbed with the polymer, hence are very likely to leach to the water and cause a toxic effect [8]. In turn, beached pellets adsorbed persistent organic pollutants, polycyclic aromatic hydrocarbons and heavy metals onto their surface at concentrations higher than those found in the environment [7]. As a result the leachate from beached pellets is likely to contain a more complex mixture of contaminants, and at higher concentrations, than the leachate from virgin pellets. This hypothesis is consistent with the observed more pronounced effects of leachates from beached than from virgin pellets. The relatively high (though locally realistic) pellet concentration considered here and the lack of information on the actual pollutants at play stress the need to assess *L. littorea* behavioural traits in response to various leachates concentrations and specific pollutants that are both

likely to be seasonal and site-dependent [12]. Note, however, that a 24-h incubation time is extremely short compared to the residence times of microplastic pellets in the ocean [1,2], which may indicate that the pollutant concentrations assessed here are not irreconcilable with *in situ* concentrations. Microplastics are also known to accumulate in intertidal gastropods [21]. Potential leachates from ingested microplastics hence warrant the need for further studies to decipher the effect of extrinsic versus intrinsic microplastic leachates. These results suggest that microplastic pellet leachates only impact the behaviour of *L. littorea* when predator cues are present. Microplastic pollution may have major implications

for marine ecosystems through direct effects on taxa that rely on chemosensory cues to escape predation. This is crucial as the effects of anthropogenic stressors on species interactions could cascade up and/or down foodwebs. *Littorina littorea* being a commercial species of gastropod across much of Europe, these findings also have potential industrial relevance.

**Data accessibility.** The dataset supporting this article has been uploaded as part of the electronic supplementary material.

**Competing interests.** I declare I have no competing interests.

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## References

- Thompson RC, Moore CJ, vom Saal FS, Swan SH. 2009 Plastics, the environment and human health: current consensus and future trends. *Phil. Trans. R. Soc. B* **364**, 2153–2166. (doi:10.1098/rstb.2009.0053)
- Barnes DKA, Galgani F, Thompson RC, Barlaz M. 2009 Accumulation and fragmentation of plastic debris in global environments. *Phil. Trans. R. Soc. B* **364**, 1985–1998. (doi:10.1098/rstb.2008.0205)
- Gall SC, Thompson RC. 2015 The impact of debris on marine life. *Mar. Pollut. Bull.* **92**, 170–179. (doi:10.1016/j.marpolbul.2014.12.041)
- Avio CG, Gorbi S, Milan M, Benedetti M, Fattorini D, d'Errico G, Pauletto M, Bargelloni L, Regoli F. 2015 Pollutants bioavailability and toxicological risk from microplastics to marine mussels. *Environ. Pollut.* **198**, 211–222. (doi:10.1016/j.envpol.2014.12.021)
- Desforges J, Galbraith M, Ross PS. 2015 Ingestion of microplastics by zooplankton in the northeast Pacific Ocean. *Arch. Environ. Contam. Toxicol.* **69**, 320–330. (doi:10.1007/s00244-015-0172-5)
- Jang M, Shim WJ, Han GM, Rani M, Song YK, Hong SH. 2016 Styrofoam debris as a source of hazardous additives for marine organisms. *Environ. Sci. Technol.* **50**, 4951–4960. (doi:10.1021/acs.est.5b05485)
- Fries E, Zarfl C. 2011 Sorption of polycyclic aromatic hydrocarbons (PAHs) to low and high density polyethylene (PE). *Environ. Sci. Pollut. Res.* **19**, 1296–1304. (doi:10.1007/s11356-011-0655-5)
- Bejgam S, MacLeod M, Bogdal C, Breitholtz M. 2015 Toxicity of leachate from weathering plastics: an exploratory screening study with *Nitocra spinipes*. *Chemosphere* **132**, 114–119. (doi:10.1016/j.chemosphere.2015.03.010)
- Law KL, Thompson RC. 2014 Microplastics in the seas. *Science* **345**, 144–145. (doi:10.1126/science.1254065)
- Hidalgo-Ruz V, Gutow L, Thompson RC, Thiel M. 2012. Microplastics in the marine environment: a review of the methods used for identification and quantification. *Environ. Sci. Technol.* **46**, 3060–3075. (doi:10.1021/es2031505)
- Holmes LA, Turner A, Thompson RC. 2014 Interactions between trace metals and plastic production pellets under estuarine conditions. *Mar. Chem.* **167**, 25–32. (doi:10.1016/j.marchem.2014.06.001)
- Gandara e Silva PP, Nobre CR, Resaffe P, Pereira CDS, Gusmao F. 2016. Leachate from microplastics impairs larval development in brown mussels. *Water Res.* **106**, 364–370. (doi:10.1016/j.watres.2016.10.016)
- Nobre CR, Santana MFM, Maluf A, Cortez FS, Cesar A, Pereira CDS, Turra A. 2015 Assessment of microplastic toxicity to embryonic development of the sea urchin *Lytechinus variegatus* (Echinodermata: Echinoidea). *Mar. Pollut. Bull.* **92**, 99–104. (doi:10.1016/j.marpolbul.2014.12.050)
- Mitchell MD, Bairos-Novak KR, Ferrari MCO. 2017 Mechanisms underlying the control of responses to predator odours in aquatic prey. *J. Exp. Biol.* **220**, 1937–1946. (doi:10.1242/jeb.135137)
- Seuront L. 2011 Hydrocarbon contamination decreases mating success in a marine planktonic copepod. *PLoS ONE* **6**, e26283. (doi:10.1371/journal.pone.0026283)
- Jacobsen HP, Stabell OB. 1999 Predator-induced alarm responses in the common periwinkle, *Littorina littorea*: dependence on season, light conditions, and chemical labelling of predators. *Mar. Biol.* **134**, 551–557. (doi:10.1007/s002270050)
- Or MV, El-Bekai M, Lui M, Watson K, Lukowiak K. 2007 Predator detection in *Lymanea stagnalis*. *J. Exp. Biol.* **210**, 4150–4158. (doi:10.1242/jeb.010173)
- Rundle SD, Spicer JI, Coleman RA, Vosper J, Soane J. 2004 Environmental calcium modifies induced defences in snails. *Proc. R. Soc. Lond. B* **271**, S67–S70. (doi:10.1098/rsbl.2003.0106)
- Bibby R, Cleall-Harding P, Rundle S, Widdicombe S, Spicer J. 2007 Ocean acidification disrupts induced defences in the intertidal gastropod *Littorina littorea*. *Biol. Lett.* **3**, 699–701. (doi:10.1098/rsbl.2007.0457)
- Cotton PA, Rundle SD, Smith KE. 2004 Trait compensation in marine gastropods: shell shape, avoidance behavior, and susceptibility to predation. *Ecology* **85**, 1581–1584. (doi:10.1890/03-3104)
- Thushari GGN, Senevirathna JDM, Yukipitiyage A, Chavanich S. 2017 Effects of microplastics on sessile invertebrates in the eastern coast of Thailand: an approach to coastal zone conservation. *Mar. Pollut. Bull.* **124**, 349–355. (doi:10.1016/j.marpolbul.2017.06.010)